

Seroprevalence of Human Herpes Simplex, Hepatitis B and Epstein-Barr Viruses in Children with Acute Lymphoblastic Leukemia in Southern Iran

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Abstract To investigate the seroprevalence of Herpes Simplex Viruses (HSV1 and HSV2), Epstein-Barr Virus (EBV) and Hepatitis B Virus (HBV) in children with acute lymphoblastic leukemia (ALL) in southern Iran. 90 patients with ALL and 90 age-sex matched healthy participants were enrolled in this study. Antibodies (IgG) against HBsAg, HSV1, HSV2, EBV different antigens including Epstein-Barr nuclear antigen-1 (EBNA-1), viral capsid antigen (VCA) and early antigen (EA) were measured by enzyme-linked immunosorbent assay (ELISA). There were 54 (60%) male and 36 (40%) female in both study groups with mean age of 8.47 ± 3.61 and 8.61 ± 2.84 years in case and control group respectively ($P=0.812$). The prevalence of antibodies against HBsAg ($P=0.002$), HSV1 ($P<0.0001$), VCA ($P=0.021$) and EA ($P<0.0001$) antigens of

EBV were significantly higher in ALL patients. With the results of this study, we could not exclude a connection between these viral infections and later leukemogenesis in childhood ALL, although nor latent infection nor congenital infection cannot be excluded by this method.

Keywords Acute Lymphoblastic Leukemia (ALL) · Epstein-Barr Virus (EBV) · Hepatitis B Virus (HBV) · Human Herpes Simplex Virus (HSV) · Iran

Introduction

Acute lymphoblastic leukemia (ALL) is the most common malignancy in children [1]. Many studies have focused on the natural history of this disease but have received little advantage. Epidemiologic studies [2, 3] suggest that infectious agents play role in at least one step in the process of ALL [4]. Defective or inhibited repair of DNA breaks will result in translocations (the hallmark genetic mutations of ALL) and this is a key step in pathogenesis of ALL. Previous studies have shown that some DNA viruses' infection, notably herpes viruses, Epstein-Barr virus (EBV) and Hepatitis B virus (HBV) interferes with cellular DNA repair mechanisms [5–7] and could lead to these characteristic genetic changes. Previous studies tried to determine the association between prenatal virus infections and development of pre-leukemic cells containing characteristic translocations; however they showed no association between infection with polyoma viruses [8], herpesviruses [9, 10], or parvovirus B19 [11] and childhood ALL. We thereby performed this study in order to investigate whether HSV, EBV and HBV infections could be correlated with ALL in southern Iran.

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Materials and Methods

In this cross-sectional case control study, a total of 90 patients were enrolled with a bone marrow biopsy-confirmed ALL admitted to Nemazee Hospital of Shiraz University of Medical Sciences, Shiraz, Iran from 2007 to 2008. The patients transfused in the last 6 months were excluded. Also, major surgery, tooth extraction, parenteral treatments, ongoing viral infections at admission to hospital, congenital abnormalities, other malignancies and serious diseases like blood culture positive sepsis, endocrine or hepatic disorders, and immune deficiency state were exclusion criteria. An age-sex matched control group of 90 healthy children was selected from those who referred to Motahari clinic (Shiraz, Iran) for routine check-ups. Those with ongoing viral infections, malignancies, congenital abnormalities, tooth extraction, previous major surgeries and parenteral therapies were excluded from the study. Written informed consent was obtained from all patients, controls and parents. The local ethics committee of the Shiraz University of Medical Sciences approved the study protocol.

A blood sample was drawn from each patient at the time of diagnosis and was transferred to Shiraz Institute for cancer Research for further analysis. For the HBV, EBV and HSV infection screening, a third-generation, sandwich enzyme-linked immunosorbent assay (ELISA) was used. All kits were purchased from Bioprobes Srl, Milan, Italy. The measurements were performed according to the manufacture's guidelines.

With this method we measured the antibodies (IgG) against HBsAg, HSV1 and HSV2 surface antigens, Epstein-Barr nuclear antigen-1 (EBNA-1), viral capsid antigen (VCA) and early antigen (EA).

Statistical Analysis

All statistical analyses were performed with the Statistical Package for Social Sciences version 15.0 (SPSS Inc., Chicago, IL, USA). The results are expressed as mean value \pm standard deviation. Chi-square test was used for comparison between groups. A two-tailed *P*-value less than 0.05 was considered statistically significant.

Results

There were 54 (60%) male and 36 (40%) female in both study groups. The mean age was 8.47 ± 3.61 and 8.61 ± 2.84 years in case and control group respectively ($P=0.812$). The seroprevalence of study viruses in both groups is shown in Table 1.

Forty four patients (48.8%) in the case group tested positive for HBs Ab while 22 (24.4%) of controls were

Table 1 The seroprevalence of HBV, EBV and HSV in patients and controls

| | Cases (<i>n</i> =90) | Controls (<i>n</i> =90) | <i>P</i> -value |
|------------------|-----------------------|--------------------------|-----------------|
| HBs Ab | 44 (48.8%) | 22 (24.4%) | 0.002 |
| EBV (EBNA-1) IgG | 40 (44.4%) | 44 (48.8%) | 0.061 |
| EBV (VCA) IgG | 28 (31.1%) | 13 (14.4%) | 0.021 |
| EBV (EA) IgG | 10 (11.1%) | 1 (1.1%) | <0.0001 |
| HSV1 IgG | 74 (82.2%) | 49 (54.4%) | <0.0001 |
| HSV2 IgG | 0 (0%) | 2 (2.2%) | 0.129 |

found to be positive ($P=0.002$). In the same way patients had higher prevalences of antibodies against VCA ($P=0.021$) and EA ($P<0.0001$) antigens of EBV as well as HSV1 ($P<0.0001$). There was no difference regarding the prevalence of EBV (EBNA-1) IgG ($P=0.061$) and HSV2 between two groups ($P=0.129$).

Discussion

In this study we investigated the relationship between the previous infection with three viruses including HSV, HBV and EBV and later ALL. These three DNA viruses have already shown to play an important role in the pathogenesis of different cancers through their interference with cellular DNA repair mechanisms [5–7]. We found that the prevalence of HBV, HSV1 and EBV was significantly higher in those who were diagnosed to have ALL. In the other hands, HSV2 and the antibodies against EBNA-1 antigen of EBV were not significantly different among cases and controls. This shows that HBV, HSV1 and EBV may play a role in the pathogenesis of ALL. However it is not known that these infections are acquired or congenital. It is hypothesized that in utero infections with potentially oncogenic viruses could be involved in pathogenesis of ALL by inducing genomic instability in B lymphocytes. Studies on twins suggest that a postnatal molecular event or exposure is required to expand pre-leukemic clone. Infectious agents especially DNA viruses can increase the stress on lymphocyte precursor proliferation and thus accelerate the transition of pre-leukemic clones to leukemic ones [12, 13].

Kinlen in two separate studies showed that the incidence of leukemia will rise in populations that have been composed of previously separated urban and rural populations by facilitating transmission of infective agents [14, 15]. Most authors use space-time clustering at diagnosis [16, 17], while others prefer space-time clustering around the time of birth [18–20]. While this may reflect exposures operating in utero, the leukemogenic infection may occur after birth since addresses used in space-time clustering studies often apply well after birth. Knox et al. have

suggested another model for virus transmission during pregnancy by observing maternal transmission of leukemia in cats [21]. This maternal transmission can be a result of either primary maternal infection or reactivation of the latent infection.

In this regard Bogdanovic and colleagues [9] used the Guthrie cards of neonates who later developed ALL to detect human herpes virus 6 (HHV-6) and EBV. But all the subjects tested negative for these two viruses. They concluded that childhood ALL is unlikely to be associated with an in utero infection with EBV or HHV-6. In the same way Gustafsson and colleagues [22] detected adenovirus DNA in 13 of 49 neonatal blood spots from ALL patients but only in 3 of 47 controls ($P=0.012$) suggesting a correlation between prenatal adenovirus infection and the development of ALL. However, in a study which was performed in Egypt, Loutfy and colleagues [23] found a higher exposure of HSV1 and HSV2 both primary infection and reactivation among ALL patients. But they couldn't correlate the ALL with EBV and CMV.

EBV has close relationship with the endemic form of Burkitt lymphoma, the most common childhood cancer in equatorial Africa and it was first discovered because of this relationship [24, 25]. Lethal lymphoproliferative disease in patients undergoing organ and stem cell transplantation is a result of EBV reactivation [19] and this demonstrates its oncological potential. Besides the transplant setting, EBV is also frequently reactivated during pregnancy [26]. Lehtinen and colleagues [27] showed that reactivation of EBV during the first trimester of pregnancy, increases the risk of ALL in the offspring.

In conclusion we showed that the seroprevalence of EBV, HSV1 and HBV was higher in patients with ALL. Hence, we could not exclude a connection between these viral infections and later leukemogenesis in childhood ALL, although nor latent infection nor congenital infection cannot be excluded by this method. In view of the epidemiological evidence for a relation between childhood ALL and infection, the search for a virus etiology must continue.

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References

- Greenlee RT, Murray T, Bolden S, Wingo PA (2000) Cancer statistics, 2000. *CA Cancer J Clin* 50:7–33
- Kinlen L (1988) Evidence for an infective cause of childhood leukemia: comparison of a Scottish new town with nuclear reprocessing sites in Britain. *Lancet* 2:1323–1327
- Kinlen L, Doll R (2004) Population mixing and childhood leukaemia: fallon and other US clusters. *Br J Cancer* 91:1–3
- Greaves M (2006) Infection, immune responses and the aetiology of childhood leukaemia. *Nat Rev Cancer* 6:193–203
- Weitzman MD, Ornelles DA (2005) Inactivating intracellular antiviral responses during adenovirus infection. *Oncogene* 24:7686–7696
- Wilkinson DE, Weller SK (2006) Herpes simplex virus type I disrupts the ATR-dependent DNA-damage response during lytic infection. *J Cell Sci* 119:2695–2703
- Sonmez M, Bektas O, Yilmaz M, Durmus A, Akdogan E, Topbas M, Erturk M, Ovali E, Omay SB (2007) The relation of lymphoma and hepatitis B virus/hepatitis C virus infections in the region of east black sea, Turkey. *Tumori* 93:536–539
- Priftakis P, Dalianis T, Carstensen J, Samuelsson U, Lewensohn-Fuchs I, Bogdanovic G, Winiarski J, Gustafsson B (2003) Human polyomavirus DNA is not detected in Guthrie cards (dried blood spots) from children who developed acute lymphoblastic leukemia. *Med Pediatr Oncol* 40:219–223
- Bogdanovic G, Jernberg AG, Priftakis P, Grillner L, Gustafsson B (2004) Human herpes virus 6 or Epstein-Barr virus were not detected in Guthrie cards from children who later developed leukaemia. *Br J Cancer* 91:913–915
- Gustafsson B, Jernberg AG, Priftakis P, Bogdanovic G (2006) No CMV DNA in Guthrie cards from children who later developed ALL. *Pediatr Hematol Oncol* 23:199–205
- Isa A, Priftakis P, Broliden K, Gustafsson B (2004) Human parvovirus B19 DNA is not detected in Guthrie cards from children who have developed acute lymphoblastic leukemia. *Pediatr Blood Cancer* 42:357–360
- Greaves M (1999) Molecular genetics, natural history and the demise of childhood leukaemia. *Eur J Cancer* 35:1941–1953
- Wiemels JL, Cazzaniga G, Daniotti M, Eden OB, Addison GM, Masera G, Saha V, Biondi A, Greaves MF (1999) Prenatal origin of acute lymphoblastic leukaemia in children. *Lancet* 354:1499–1503
- Kinlen L, Jiang J, Hemminki K (2002) A case-control study of childhood leukaemia and paternal occupational contact level in rural Sweden. *Br J Cancer* 86:732–737
- Kinlen LJ, Balkwill A (2001) Infective cause of childhood leukaemia and wartime population mixing in Orkney and Shetland. *Lancet* 358:155
- Petridou E, Revinthi K, Alexander FE, Haidas S, Kolioukas D, Kosmidis H, Piperopoulos F, Tzortzatos F, Trichopoulos D (1996) Space-time clustering of childhood leukaemia in Greece: evidence supporting a viral aetiology. *Br J Cancer* 73:1278–1283
- Gilman EA, McNally RJ, Cartwright RA (1999) Space-time clustering of acute lymphoblastic leukaemia in parts of the U.K. (1984–1993). *Eur J Cancer* 35:91–96
- Gustafsson B, Carstensen J (1999) Evidence of space-time clustering of childhood acute lymphoblastic leukaemia in Sweden. *Br J Cancer* 79:655–657
- Gustafsson B, Carstensen J (2000) Space-time clustering of childhood lymphatic leukaemia and non-Hodgkin's lymphomas in Sweden. *Eur J Epidemiol* 16:1111–1116
- Birch JM, Alexander FE, Blair V, Eden OB, Taylor GM, McNally RJ (2000) Space-time clustering patterns in childhood leukaemia support a role for infection. *Br J Cancer* 82:1571–1576
- Knox EG, Stewart A, Kneale G (1980) Childhood leukaemia and mother—foetus infection. *Br J Cancer* 42:158–161
- Gustafsson B, Huang W, Bogdanovic G, Gauffin F, Nordgren A, Talekar G, Ornelles DA, Gooding LR (2007) Adenovirus DNA is detected at increased frequency in Guthrie cards from children who develop acute lymphoblastic leukaemia. *Br J Cancer* 97:992–994

23. Loutfy SA (2006) HM, Ibrahim MF, Hafez MM. Seroprevalence of herpes simplex virus types 1 and 2, Epstein-Barr virus, and cytomegalovirus in children with acute lymphoblastic leukemia in Egypt. *Saudi Med J* 27:1139–1145
24. Burkitt D (1958) A sarcoma involving the jaws in African children. *Br J Surg* 46:218–223
25. Epstein MA, Achong BG, Barr YM (1964) Virus particles in cultured lymphoblasts from Burkitt's lymphoma. *Lancet* 15:702–703
26. Costa S, Barrasso R, Terzano P, Zerbini M, Carpi C, Musiani M (1985) Detection of active Epstein–Barr infection in pregnant women. *Eur J Clin Microbiol* 4:335–336
27. Lehtinen M, Koskela P, Ogmundsdottir HM, Bloigu A, Dillner J, Gudnadottir M, Hakulinen T, Kjartansdottir A, Kvarnung M, Pukkala E, Tulinius H, Lehtinen T (2003) Maternal herpesvirus infections and risk of acute lymphoblastic leukemia in the offspring. *Am J Epidemiol* 158:207–213