

# Loss of MTSS1 Expression is an Independent Prognostic Factor for Hilar Cholangiocarcinoma

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**Abstract** Metastasis suppressor 1 (MTSS1) is a novel metastasis suppressor gene in a variety of cancers. This study aimed to detect MTSS1 expression in normal and cancerous tissue specimens from Chinese patients with hilar cholangiocarcinoma to determine the association with clinicopathological parameters and survival. Tissue microarrays containing normal and tumor specimens were constructed using paraffin blocks from 61 patients for immunohistochemical analysis of MTSS1 expression. A subgroup of these tissues was verified by Western blot analysis. MTSS1 protein was expressed in 24 of 61 cases (39.3 %) of tumor tissues, compared to that in 22 of 26 (84.6 %) of non-neoplastic bile duct epithelium and in 26 of 26 (100 %) of adjacent normal liver cells. Loss of MTSS1 expression was associated with lymph node metastases of cholangiocellular carcinoma and tumor cell de-differentiation. MTSS1 expression inversely associated with tumor recurrence and overall survival of the patients by univariate and multivariate analyses. MTSS1 expression was significantly decreased in human hilar cholangiocarcinoma and lost MTSS1 expression was associated with poor overall survival and tumor recurrences in cholangiocarcinoma patients. Thus, MTSS1 expression represents an independent predictor for tumor recurrence and overall survival in patients with cholangiocarcinoma.

**Keywords** Cholangiocarcinoma · MTSS1 · Immunohistochemistry · Prognosis

## Introduction

Hilar cholangiocarcinoma is an extremely lethal tumor with overall five-year survival rates of less than 20 %. Due to the location at the biliary tree, this deadly disease is usually diagnosed at later stages, which precludes the effectiveness of conventional therapies [1–3]. However, early stage hilar cholangiocarcinoma can be easily treated by R0 resection with negative histological margins, and this intervention improves the five-year survival rate to 30–50 % [1, 2, 4–6]. The etiology and pathogenesis of cholangiocarcinoma remain to be defined, and a number of pathologic conditions resulting in either acute or chronic biliary tract epithelial injury may predispose an individual to malignant transformation. Thus, more research on hilar cholangiocarcinoma is urgently needed for improvement of early diagnosis, initiation and development of more effective treatment strategies, and better prediction of the patient outcome.

The metastasis suppressor 1 (MTSS1) gene, mapped to human chromosome 8q24.1, is expressed in non-metastatic tumor cell lines, but remarkably absent in metastatic or invasive cancer cells [7–10]. Previous studies indicated that MTSS1 protein might play a critical role in regulating cytoskeletal dynamics, cell proliferation, and carcinogenesis in various organ sites (7–13). For example, MTSS1 expression has been clinically detected as dramatically decreased in cancers of the prostate [7], bladder [7, 9], stomach [11] and mammary glands [10, 12]. Moreover, decreased MTSS1 was found to be associated with tumor progression and poor survival (9–11), which suggests a potential role in suppressing cancer development. Loss of expression of MTSS1 protein was shown to be induced by DNA methylation of its gene promoter (8). Nevertheless, other studies have shown that MTSS1 was highly expressed in hepatocellular carcinoma [13]. The MTSS1 protein has been characterized as a sonic

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hedgehog-responsive gene that potentiates Gli-dependent transcription; MTSS1 appears to regulate target gene expression through its association with the Gli complex, which is responsible for epidermal growth [14].

To date, there are no data available about MTSS1 expression in cholangiocarcinoma. The aims of this study were to detect MTSS1 expression in a large number of hilar cholangiocarcinoma tissue specimens and to investigate the potential association of MTSS1 expression with clinicopathological features and survival of hilar cholangiocarcinoma patients. Our data showed the loss of MTSS1 expression might serve as a useful biomarker in predicting the outcome of patients with cholangiocarcinoma.

## Material and Methods

### Tissue Samples and Tissue Microarray Construction

Sixty-one patients with primary hilar cholangiocarcinoma were identified for study participation from Eastern Hepatobiliary Hospital and Changzheng Hospital, Shanghai, China between 2005 and 2007. Haematoxylin-eosin (HE)-stained sections of resected tumor samples were prepared and reviewed by two pathologists to ensure the quality of paraffin-embedded tissue blocks. Medical records of these patients were reviewed for extraction of clinicopathological data, including age at diagnosis, sex, tumor size, depth of invasion, presence of nodal metastases, and the American Joint Committee on Cancer stage. The mean age of patients at tumor resection was 55 years old (range: 31–79 years old). There were 45 (73.8 %) males and 16 (26.2 %) females. None of these patients received any pre-surgery treatment. The median follow-up period of time for these patients was 16 months (range: 1–59 months). Three patients were lost during follow-up and four patients died of other diseases without recurrence [15]. The Institutional Review Board of both Eastern Hepatobiliary Hospital and Changzheng Hospital approved the use of the tissues and clinical information and informed consent was obtained from each patient or their guardians.

Tissue blocks (tumor and patient-matched normal tissue specimens) were retrieved from the respective Pathology Departments and used to construct the two paraffin-embedded tissue microarray (TMA) blocks using a manual arrayer (Beecher Instruments, Sun Prairie, WI, USA) as previously described [15].

### Protein Extraction and Western Blot Analysis

Whole-cell lysates were prepared from both normal and cancerous tissue specimens using a sodium dodecyl sulfate (SDS) lysis buffer. After quantification of the isolated protein, each patient's sample was separated in SDS-polyacrylamide gel

electrophoresis gels and then detected by standard Western blotting technique. The primary antibody was rabbit polyclonal antibody against human MTSS1 (1:500 dilution; Abnova, Buckingham, UK) and the secondary antibody was horseradish peroxidase-linked donkey anti-rabbit IgG antibody (1:5000; Amersham, Arlington Heights, IL, USA). The anti- $\beta$ -actin antibody was used as an internal control to normalize sample loading. The targeted protein bands were detected by an enhanced chemiluminescence kit from Amersham, according to the manufacturer's instructions.

### Immunohistochemistry

Four-micron paraffin-embedded sections from the tissue microarrays were prepared as described previously [15], and immunohistochemical staining of MTSS1 protein was conducted using a rabbit polyclonal anti-MTSS1 antibody (1:50; Abnova, Caltag-MedSystems Ltd., Buckingham, UK). A Streptavidin-Biotin kit (#KIT-9720; Maixin, Fuzhou, China) was used to visualize antibody binding to the tissues. Counterstaining was performed with haematoxylin. Control sections were incubated with buffer only, instead of the primary antibody.

### Review and Scoring of Immunohistochemically-Stained Sections

The stained TMA sections were independently evaluated and scored under a microscope by two pathologists without any knowledge of clinical outcome. Any discrepancy was dissolved by re-evaluation of the sections by these two pathologists. A semi-quantitative scoring system was used to assess MTSS1 expression, as described previously [16]. Briefly, MTSS1 staining was graded on a binary scale of 0 or 1 (0, negative staining [ $<10$  % of tumor cells stained cytoplasmically]; 1, positive staining [ $\geq 10$  % of tumor cells stained cytoplasmically]) as described previously [17] (Figure S1).

### Statistical Analyses

The data were summarized as high and low immunohistochemical expression of MTSS1 protein and then statistically analyzed using Chi-square test. Within-group correlations of continuous and ordinal variables were assessed using Pearson's correlation coefficient or Spearman's rank correlation coefficient. The Kaplan-Meier method was used to estimate survival rates, and the log-rank and the Wilcoxon rank sum tests were performed to assess survival differences between groups. The Cox proportional hazards model for multivariate survival analysis was used to assess predictors related to survival. A two-sided *P*-value of less than 0.05 was considered statistically significant. Analyses were

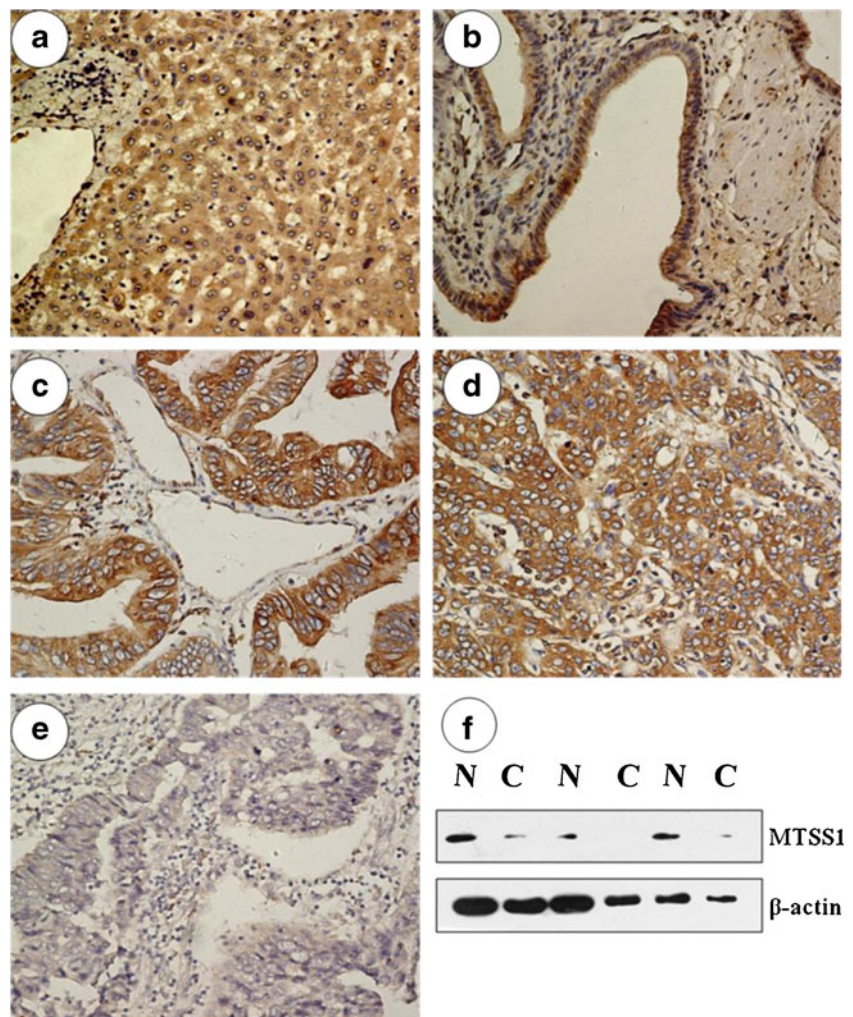
performed using the SPSS statistical software program for Microsoft Windows (SPSS Inc., Chicago, IL, USA).

## Results

### Differential Expression of MTSS1 Protein in Cholangiocellular Carcinoma and Adjacent Normal Tissues

Data from both immunohistochemistry and Western blotting showed that all cholangiocytes of normal bile ducts and the normal hepatocytes had moderate or strong staining of MTSS1 (Fig. 1a and b). In contrast, MTSS1 expression was significantly decreased in tumor cells of cholangiocellular carcinoma, as compared to that in adjacent normal tissues (Fig. 1c–f). Particularly, MTSS1 expression was observed in 22 of 26 (84.6 %) non-neoplastic bile duct epithelium and 26 of 26 (100 %) adjacent normal liver cells. MTSS1 expression was only found in 24 of 61 (39.3 %) cases of primary hilarcholangiocellular carcinoma, significantly lower than that in para-neoplastic normal bile ducts ( $P < 0.001$ ).

**Fig. 1** Expression of MTSS1 in hilarcholangiocarcinoma and non-neoplastic bile ducts by immunohistochemistry (a–e) and Western blotting (f). **a** Adjacent normal liver tissues; **b** Normal (non-neoplastic) bile ducts; **c, d** Positive expression; **e** Negative expression. Original magnification: 200 $\times$ . MTSS1 expression was decreased in cancer (C) but not in normal tissues (N)



### Loss of MTSS1 Expression Associated with Tumor Lymph Node Metastasis and Pathology Grade

To delineate the clinical significance of the loss of MTSS1 expression, we associated it with clinicopathological parameters from hilarcholangiocarcinoma patients. In particular, MTSS1 expression was found to be more frequently lost in tumors with regional lymph nodemetastasis (25.0 %) than in N0-stage tumors (55.2 %;  $P = 0.003$ ). Moreover, MTSS1 expression was significantly higher in well- to moderately-differentiated tumors than in low- to un-differentiated tumors (55.9 % vs. 18.5 %;  $P < 0.008$ ). However, there was no significant association between MTSS1 expression and other clinicopathological factors, such as tumor size, nerve invasion, tumor invasion, and clinical stage Table 1.

### Association of MTSS1 Expression with Prognosis and Recurrence of Hilarcholangiocarcinoma Patients

The median cumulative survival duration in these patients was 16 months, ranging from 1 to 59 months. MTSS1 expression

**Table 1** Association of MTSS1 expression with clinicopathological parameters of hilarcholangiocarcinoma patients

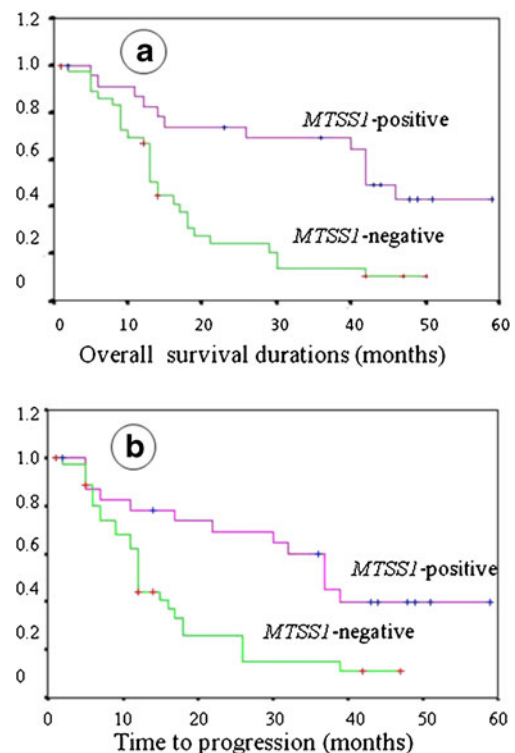
	No. of patients (%)	MTSS1 expression (%)	<i>P</i> -value
<b>Age, years</b>			
≤55	32(52.5)	14(43.8)	0.459
>55	29(47.5)	10(34.5)	
<b>Gender</b>			
Male	45 (73.8)	18(40.0)	0.860
Female	16 (26.2)	6(37.5)	
<b>Clinical stage</b>			
I–III	13(21.3)	7(53.8)	0.338
IV	48(78.7)	17(35.4)	
<b>Tumor stage,pT</b>			
T1/T2	13(21.3)	7(53.8)	0.338
T3/T4	48(78.7)	17(35.4)	
<b>Tumor differentiation</b>			
Well/Moderate	34(55.7)	19(55.9)	0.003
Poor	27(44.3)	5(18.5)	
<b>Lymph nodemetastasis,pN</b>			
No	29(47.5)	16(55.2)	0.016
Yes	32(52.3)	8(25.0)	
<b>Nerve invasion</b>			
No	34(55.7)	13(38.6)	0.842
Yes	27(44.3)	11(40.7)	
<b>Tumor size, cm</b>			
<3	22(36.1)	6(27.3)	0.147
≥3	39(63.9)	18(46.2)	
Total cases	61	24(39.3)	

in the tumor tissues was associated with longer disease-free survival of the patients. Specifically, patients with MTSS1-negative tumors had a median survival of 14 months, whereas patients with MTSS1-positive tumors had a median survival of 42 months ( $P=0.005$ ) (Fig. 2a). The other factors in univariate analysis were tumor invasion ( $P=0.0085$ ), lymph node metastasis ( $P=0.0012$ ), residual tumor margin (R1 or R2) ( $P=0.0295$ ), tumor differentiation ( $P=0.0027$ ), and TNM stage ( $P=0.0085$ ). Multivariate analysis by Cox regression revealed that complete resection ( $P=0.045$ ), lymph node metastasis ( $P=0.015$ ) and MTSS1 ( $P=0.021$ ) were independent prognostic factors. Furthermore, patients with MTSS1-positive tumors were associated with a future recurrence at an average of 37 months, whereas patients with MTSS1-negative tumors had recurrence at an average of 12 months ( $P=0.0025$ ) (Fig. 2b). Other factors that associated with future recurrence in univariate analysis were tumor invasion ( $P=0.0039$ ), residual tumor margin (R1 or R2) ( $P=0.022$ ), tumor cell differentiation ( $P=0.0008$ ), lymph node metastasis ( $P=0.0005$ ), and TNM stage ( $P=0.0039$ ). Multivariate analysis using the Cox proportional hazards model for all significant variables in univariate analysis showed that tumor invasion ( $P=0.009$ ), lymph node metastasis

( $P=0.029$ ), and MTSS1 ( $P=0.021$ ) were all independent predictors for tumor recurrence.

## Discussion

In the present study, we enrolled 61 patients with cholangiocellular carcinoma, collected their clinicopathological data, and constructed tissue microarrays using the paraffin blocks obtained from these patients. We then performed immunohistochemical staining of MTSS1 protein in both normal and hilarcholangiocarcinoma tissue specimens. Our data showed that MTSS1 expression was significantly reduced in hilarcholangiocarcinoma specimens, as compared to the patient-matched normal tissue specimens. Further statistical analyses showed that the loss of MTSS1 expression was associated with lymph node metastases of cholangiocellular carcinoma and tumor cell de-differentiation. MTSS1 expression was inversely associated with overall survival of the patients and tumor recurrence. The univariate and multivariate analyses revealed that detection of MTSS1 expression is an independent predictor for both overall survival of the



**Fig. 2** Kaplan-Meier analysis of overall survival of and recurrence in hilarcholangiocarcinoma patients according to MTSS1 expression. **a** Overall survival was significantly worse in patients with MTSS1-negative tumors (median survival, 14 months) than in those with MTSS1-positive tumors (median survival, 42 months;  $P<0.001$ ). **b** Disease-free survival was significantly worse in patients with MTSS1-negative tumors (median survival, 12 months) than in those with MTSS1-positive tumors (median survival, 37 months;  $P<0.001$ )

patients and future recurrence. Future studies with larger patient populations will be carried out to confirm this finding.

Cholangiocarcinoma is a relatively rare disease, accounting for only about 2 % of all cancers diagnosed worldwide. The long-term survival of the patients remains dismal, although great advances have been made in therapeutic strategies available to treat this cancer at early stages. Numerous clinicopathological factors that affect clinical outcome of hilarcholangiocarcinoma have been investigated, including TNM stage, curative resection, and lymph node metastasis of the tumor [1, 2, 4–6, 18, 19]. Of these, complete resection (R0 resection) has been the most frequently used factor to predict long-term survival [19, 20]. Data from our previous studies have also shown that those patients with extensive tumor invasion and lymph node metastasis survive for a much shorter period and experience higher rates of recurrence than those with localized tumors and less invasion [21, 22]. The data from this study indicated that lymph node metastasis and curative resection were independent factors for the long-term survival of patients with hilarcholangiocarcinoma. Moreover, R0 resection was able to achieve long-term survival, as compared to resections with macroscopic- and microscopic-positive margins. In addition, TNM stage did not seem to be an independent predictor for overall survival or tumor recurrence in these patients.

Molecular events underlying the development of hilarcholangiocarcinoma are gradually being elucidated, and a number of studies have provided information on gene expression patterns that may prove useful as biomarkers to predict the overall survival in patients after resection of hilarcholangiocarcinoma. For example, previous studies have shown that differential gene expressions of MMP [23], p27 [24], MVD [25], or Anxa1 [15] can distinguish tumors from normal tissues and these genes' expressions can be used for early tumor detection and survival prediction. In contrast, other studies have failed to find a correlation between some of these markers and prognosis of the patients [26]. In our current study, we detected, for the first time, differential MTSS1 expression in hilarcholangiocarcinoma and the corresponding normal tissues, suggesting this gene's potential as a novel tumor biomarker. Indeed, previous studies of other types of cancers have demonstrated the usefulness of MTSS1 protein as a biomarker [7, 9, 11, 12]. Our current data showed that MTSS1 expression was significantly reduced in hilarcholangiocarcinoma tissues, as compared to the normal tissues. The loss of expression of MTSS1 protein was associated with lymph node metastasis and tumor de-differentiation, but was not associated with tumor stage and size, nerve invasion, or tumor invasion. This result was in agreement with previous studies in other cancers, including those of the stomach and breast. Moreover, the loss of MTSS1 expression was associated with shorter overall survival of the patients and with more frequent recurrence. Multivariate analysis confirmed this finding and identified MTSS1 as an independent

factor for predicting future recurrence and overall survival of patients with cholangiocarcinoma. However, future study will be needed to independently verify these findings.

Our current data indicate that MTSS1 protein may play an important role in suppression of hilarcholangiocarcinoma development and progression, although the functions of MTSS1 protein in hilarcholangiocarcinoma are still not fully elucidated. Molecularly, MTSS1 contains a C-terminal WH2 (WASP homology domain-2) domain, which is a small actin monomer-binding motif that interacts with Rac, an actin and actin-associated protein, to modulate lamellipodia formation during cell migration [27–30]. Enhanced expression of MTSS1 has been demonstrated to suppress the invasive, migration, growth and adherence properties of breast cancer cell lines, and knockdown of MTSS1 has been shown to dramatically enhance these properties [10]. Further studies are needed to explore the functions and the underlying mechanism of MTSS1 in hilarcholangiocarcinoma.

It is important to note that our current study has some limitations. First, most of the patients used in this study were diagnosed with stage IV disease, which may have introduced some bias in the analysis and results. Second, the study is associative and was carried out without any mechanistic considerations. Therefore, future study with a larger sample size and more diversified tumor stages will be needed to verify our current findings before clinical use of MTSS1 as tumor marker.

In conclusion, the present study demonstrated that loss of MTSS1 expression was a common event in hilarcholangiocarcinoma and associated with lymph node metastasis and poor differentiation of hilarcholangiocarcinoma. After independent confirmation of these findings is achieved, detection of MTSS1 expression could be useful as a tumor biomarker in early detection and survival prediction for patients with hilarcholangiocarcinoma.

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## References

- Ito F, Cho CS, Rikkers LF, Weber SM (2009) Hilar cholangiocarcinoma: Current management. *Ann Surg* 250(2):210–218
- Otani K, Chijiwa K, Kai M, Ohuchida J, Nagano M, Tsuchiya K, Kondo K (2008) Outcome of surgical treatment of hilar cholangiocarcinoma. *J Gastrointest Surg* 12(6):1033–1040

3. Liu CL, Fan ST, Lo CM, Tso WK, Lam CM, Wong J (2006) Improved operative and survival outcomes of surgical treatment for hilar cholangiocarcinoma. *Br J Surg* 93(12):1488–1494
4. Sawada T, Kita J, Rokkaku K, Kato M, Shimoda M, Kubota K (2008) Outcome of surgical resection for hilar cholangiocarcinoma in elderly patients. *Hepatogastroenterology* 55(88):1971–1974
5. Seyama Y, Makuuchi M (2007) Current surgical treatment for bile duct cancer. *World J Gastroenterol* 13(10):1505–1515
6. Maeno H, Ono T, Yamanoi A, Nagasue N (2007) Our experiences in surgical treatment for hilar cholangiocarcinoma. *Hepatogastroenterology* 54(75):669–673
7. Nixdorf S, Grimm MO, Loberg R, Marreiros A, Russell PJ, Pienta KJ, Jackson P (2004) Expression and regulation of MIM (Missing In Metastasis), a novel putative metastasis suppressor gene, and MIM-B, in bladder cancer cell lines. *Cancer Lett* 215(2):209–220
8. Utikal J, Gratchev A, Muller-Molinat I, Oerther S, Kzhyshkowska J, Arens N, Grobholz R, Kannookadan S, Goerdts S (2006) The expression of metastasis suppressor MIM/MTSS1 is regulated by DNA methylation. *Int J Cancer* 119(10):2287–2293
9. Wang Y, Liu J, Smith E, Zhou K, Liao J, Yang GY, Tan M, Zhan X (2007) Downregulation of missing in metastasis gene (MIM) is associated with the progression of bladder transitional carcinomas. *Cancer Invest* 25(2):79–86
10. Parr C, Jiang WG (2009) Metastasis suppressor 1 (MTSS1) demonstrates prognostic value and anti-metastatic properties in breast cancer. *Eur J Cancer* 45(9):1673–1683
11. Liu K, Wang G, Ding H, Chen Y, Yu G, Wang J (2007) Downregulation of metastasis suppressor 1 (MTSS1) is associated with nodal metastasis and poor outcome in Chinese patients with gastric cancer. *BMC Cancer* 10:428
12. Hicks DG, Yoder BJ, Short S, Tarr S, Prescott N, Crowe JP, Dawson AE, Budd GT, Sizemore S, Cicek M et al (2006) Loss of breast cancer metastasis suppressor 1 protein expression predicts reduced disease-free survival in subsets of breast cancer patients. *Clin Cancer Res* 12(22):6702–6708
13. Ma S, Guan XY, Lee TK, Chan KW (2007) Clinicopathological significance of missing in metastasis B expression in hepatocellular carcinoma. *Hum Pathol* 38(8):1201–1206
14. Callahan CA, Ofstad T, Hornig L, Wang JK, Zhen HH, Coulombe PA, Oro AE (2004) MIM/BEG4, a Sonic hedgehog-responsive gene that potentiates Gli-dependent transcription. *Genes Dev* 18(22):2724–2729
15. Wang D, Zhang H, Fang Z, Yu G (2007) Annexin-1 downregulation is associated with clinical outcome in Chinese patients with hilar cholangiocarcinoma. *Eur Surg Res* 45(3–4):151–157
16. Liu JH, Song LB, Zhang X, Guo BH, Feng Y, Li XX, Liao WT, Zeng MS, Huang KH (2008) Bmi-1 expression predicts prognosis for patients with gastric carcinoma. *J Surg Oncol* 97(3):267–272
17. Urano N, Fujiwara Y, Doki Y, Tsujie M, Yamamoto H, Miyata H, Takiguchi S, Yasuda T, Yano M, Monden M (2006) Overexpression of hypoxia-inducible factor-1 alpha in gastric adenocarcinoma. *Gastric Cancer* 9(1):44–49
18. Tsai HM, Chuang CH, Lin XZ, Chen CY (2009) Factors relating to the short term effectiveness of percutaneous biliary drainage for hilar cholangiocarcinoma. *World J Gastroenterol* 15(41):5206–5210
19. Tsalis K, Vasiliadis K, Kalpakidis V, Christoforidis E, Avgerinos A, Botsios D, Megalopoulos A, Haidich AB, Betsis D (2007) A single-center experience in the management of Altemeier-Klatskin tumors. *J Gastrointest Liver Dis* 16(4):383–389
20. Ramacciato G, Nigri G, Bellagamba R, Petrucciani N, Ravaioli M, Cescon M, Del Gaudio M, Ercolani G, Di Benedetto F, Cautero N et al (2010) Univariate and multivariate analysis of prognostic factors in the surgical treatment of hilar cholangiocarcinoma. *Am Surg* 76(11):1260–1268
21. Murakami Y, Uemura K, Sudo T, Hashimoto Y, Nakashima A, Kondo N, Sakabe R, Ohge H, Sueda T (2011) Prognostic factors after surgical resection for intrahepatic, hilar, and distal cholangiocarcinoma. *Ann Surg Oncol* 18(3):651–658
22. Li Q, Li HK, Hao XS (2009) Analysis of the surgical outcome and prognostic factors for hilar cholangiocarcinoma. *Zhonghua Wai Ke Za Zhi* 47(2):94–97
23. Kirimlioglu H, Turkmen I, Bassullu N, Dirican A, Karadag N, Kirimlioglu V (2009) The expression of matrix metalloproteinases in intrahepatic cholangiocarcinoma, hilar (Klatskin tumor), middle and distal extrahepatic cholangiocarcinoma, gallbladder cancer, and ampullary carcinoma: role of matrix metalloproteinases in tumor progression and prognosis. *Turk J Gastroenterol* 20(1):41–47
24. Fiorentino M, Altimari A, D'Errico A, Gabusi E, Chieco P, Masetti M, Grigioni WF (2001) Low p27 expression is an independent predictor of survival for patients with either hilar or peripheral intrahepatic cholangiocarcinoma. *Clin Cancer Res* 7(12):3994–3999
25. Thelen A, Scholz A, Benckert C, Schroder M, Weichert W, Wiedenmann B, Neuhaus P, Jonas S (2008) Microvessel density correlates with lymph node metastases and prognosis in hilar cholangiocarcinoma. *J Gastroenterol* 43(12):959–966
26. Nishihara Y, Aishima S, Hayashi A, Iguchi T, Fujita N, Taketomi A, Honda H, Tsuneyoshi M (2009) CD10+ fibroblasts are more involved in the progression of hilar/extrahepatic cholangiocarcinoma than of peripheral intrahepatic cholangiocarcinoma. *Histopathology* 55(4):423–431
27. Bershteyn M, Atwood SX, Woo WM, Li M, Oro AE (2005) MIM and cortactin antagonism regulates ciliogenesis and hedgehog signaling. *Dev Cell* 19(2):270–283.
28. Bompard G, Sharp SJ, Freiss G, Machesky LM (2005) Involvement of Rac in actin cytoskeleton rearrangements induced by MIM-B. *J Cell Sci* 118(Pt 22):5393–5403
29. Lin J, Liu J, Wang Y, Zhu J, Zhou K, Smith N, Zhan X (2005) Differential regulation of cortactin and N-WASP-mediated actin polymerization by missing in metastasis (MIM) protein. *Oncogene* 24(12):2059–2066
30. Mattila PK, Pykalainen A, Saarikangas J, Paavilainen VO, Vihinen H, Jokitalo E, Lappalainen P (2007) Missing-in-metastasis and IRSp53 deform PI(4,5)P2-rich membranes by an inverse BAR domain-like mechanism. *J Cell Biol* 176(7):953–964